ISOLATION AND STRUCTURAL ELUCIDATION OF SEKOTHRIXIDE, A NEW MACROLIDE EFFECTIVE TO OVERCOME DRUG-RESISTANCE OF CANCER CELL

Sir:

The development of multidrug-resistance (MDR) is a major limiting factor to successful chemotherapy of cancer in humans. MDR is a form of drug-resistance characterized by decreased cellular sensitivity to a broad range of chemotherapeutic agents. This resistance is due at least in part to the mdr1 gene product, P-glycoprotein, and results in the failure of chemotherapeutic treatment¹).

During the screening for new pharmacologically active agents which will overcome MDR with colchicine-resistant KB cell line $(KB-C2)^{2}$ we isolated a new antibiotic stipiamide from *Myxococcus stipitatus*³. In this paper, we report the isolation and structural elucidation of a new macrolide, named sekothrixide, which exhibits cytocidal activity against KB-C2 in the presence of colchicine $(1.5 \,\mu g/ml)$.

The producing microorganism, *Saccharothrixide* sp. CF24, was isolated from a soil sample collected at Yokose, Saitama, Japan[†].

Fermentation was carried out for 3 days at 27°C in a 50-liter jar fermenter containing 30 liters of medium with agitation of 400 rpm and aeration of 30 liters/minute. The medium consisted of soluble starch 2.5%, soybean meal 1.5%, dry yeast 0.2% and CaCO₃ 0.4% (pH 6.2 before sterilization). The mycelium cake, harvested by centrifugation from the whole fermented broth, was stirred with 5 liters of acetone. The solvent extract was concd in vacuo to a small volume and the active material was extracted from the aqueous residue twice with 500 ml each of EtOAc. The separated organic layer was dried over Na_2SO_4 and concd in vacuo. The oily residue was applied to a column of silica gel (Wakogel C-200, 6×36 cm) packed with CHCl₃ and the column was developed with CHCl₃-MeOH (150:1). Active fractions were combined and concentrated to give a brownish oil. Further purification of the oil was achieved by silica gel preparative TLC (Merck, Kieselgel 60F₂₅₄, 0.5 mm) developed with CHCl₃ MeOH (19:1) followed by reversed phase chromatography (SEP-PAK C18) developed with MeOH- H_2O (87:13) to yield 5 mg of sekothrixide as a

colorless oil (wax appearance, mp 40.5° C). Its physico-chemical properties are summarized in Table 1.

Table 1. Physico-chemical properties of sekothrixide.

Appearance	Colorless oil
MP	40.5°C
Molecular formula	$C_{25}H_{50}O_{6}$
Solubility	
Soluble in	Benzene, CHCl ₃ , MeOH
Insoluble in	Hexane, H ₂ O
Rf value	0.75 (CHCl ₃ - MeOH, 19:1)
(Silica gel 60)	0.71 (Hexane-EtOAc, 2:3)
HRFAB-MS (m/z)	$483.3665 (M + H)^+,$
	Calcd: 483.3686
UV λ_{\max}^{MeOH} nm (ε)	207 (6,370)
$\hat{\lambda}_{\max}^{MeOH + NaOH} nm (\varepsilon)$	207 (20,000), 276 (5,450)
IR v_{max} (CHCl ₃) cm ⁻¹	3460, 1729, 1706
$[\alpha]_{D}^{22}$ (c 1.0, MeOH)	-45.13°

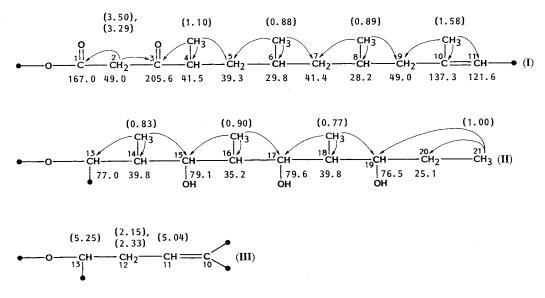
Table 2. ¹³C and ¹H NMR signals of sekothrixide taken in CDCl₃.

No.	$\delta_{\rm C}$	$\delta_{ m H}$
1	167.0	
2	49.0	3.50 (1H, d, J = 13 Hz),
		3.29 (1H, d, J = 13 Hz)
3	205.6	
4	41.5	2.91 (1H, m)
5	39.3	0.94 (1H, m), 1.88 (1H, m)
6	29.8	1.55 (1H, m)
7	41.4	0.51 (1H, ddd, $J = 7$ Hz),
		1.30 (1H, ddd, $J = 7 \text{ Hz}$)
8	28.2	1.61 (1H, m)
9	49.0	2.00 (1H, dd, $J = 6$, 13 Hz),
		1.69 (1H, dd, $J=6$, 13 Hz)
10	137.3	
11	121.6	5.04 (1H, dd, J = 7 Hz)
12	27.1	2.15 (1H, br d, $J = 13$ Hz),
		2.33 (1H, m)
13	77.0	5.25 (1H, d, $J = 11$ Hz)
14	39.8	2.03 (1H, m)
15	79.1	3.62 (1H, d, J = 10 Hz)
16	35.2	1.67 (1H, m)
17	79.6	3.84 (1H, d, J = 10 Hz)
18	39.8	1.88 (1H, m)
19	76.5	3.72 (1H, br s)
20	25.1	1.53 (2H, m)
21	11.1	1.00 (3H, t, $J = 7.5$ Hz)
4-CH ₃	19.3	1.10 (3H, d, $J = 7$ Hz)
6-CH ₃	23.3	0.88 (3H, d, $J = 7.5$ Hz)
8-CH ₃	21.1	0.89 (3H, d, J = 6 Hz)
10-CH ₃	16.6	1.58 (3H, s)
14-CH ₃	10.8	0.83 (3H, d, $J = 7.5$ Hz)
16-CH ₃	3.9	0.90 (3H, d, J = 7.5 Hz)
18-CH ₃	12.0	0.77 (3H, d, J=7.5 Hz)

[†] The paper pertaining to the taxonomy and isolation of this strain will be published elsewhere.

Fig. 1. Partial structures of sekothrixide revealed by HMBC (I and II) and ¹H-¹H COSY (III).

Values and those in parentheses show ¹³C- and ¹H-chemical shifts in ppm, respectively. Arrows mean long range ¹³C-¹H couplings.



The molecular formula of sekothrixide was determined to be $C_{28}H_{50}O_6$ by HRFAB-MS data $(m/z \ 483.3665 \ (M + H)^+$, Calcd: 483.3686). The ¹³C and ¹H NMR spectra data of sekothrixide are tabulated in Table 2. The signals in the ¹³C NMR spectrum at $\delta_C \ 167.0$ and 205.6 were assigned to an ester and a ketone due to IR absorption bands at 1729 and 1706 cm⁻¹, respectively.

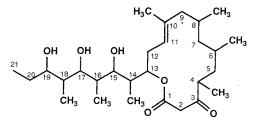
Functional groups in sekothrixide were classified as $CH_3 \times 8$, $CH_2 \times 6$, $CH \times 6$, $CH-O \times 4$, $CH=\times 1$, $C=\times 1$, $COO \times 1$ and $CO \times 1$. These accounted for three of the four degrees of unsaturation required by the molecular formula indicating sekothrixide to be monocyclic.

Detailed NMR spectral analysis of sekothrixide was aided by the use of heteronuclear multiple-bond correlation (HMBC)^{4,5)}, since it is a particularly effective method for the structural elucidation of complicated molecules with many methyl groups⁶). The partial structure I in Fig. 1 was determined through the analysis of ¹H-¹³C long range couplings observed with methyl groups. A methyl proton doublet at $\delta_{\rm H}$ 1.10 (4-CH₃) was coupled to a ketone ($\delta_{\rm C}$ 205.6, C-3), a non-oxygenated methine ($\delta_{\rm C}$ 41.5, C-4) and a methylene ($\delta_{\rm C}$ 39.3, C-5). Another methyl doublet at $\delta_{\rm H}$ 0.88 (6-CH₃) was coupled to a methylene ($\delta_{\rm C}$ 41.4, C-7), a methine ($\delta_{\rm C}$ 29.8, C-6) and the methylene just explained (C-5). These results provided the connectivity of -C3(=O)-C4H(CH₃)- $C5H_2-C6H(CH_3)-C7H_2-$. By repeating the same procedure, this partial connectivity was extended to C-11. In addition, non-equivalent methylene proton doublets at $\delta_{\rm H}$ 3.50 and 3.29 (2-H) showed cross peaks with an ester carbonyl signal at $\delta_{\rm C}$ 167.0 (C-1) and the ketone signal at $\delta_{\rm C}$ 205.6 (C-3). The partial structure thus established as shown in Fig. 1 (I) contained a β -ketoester group, whose system was supported by the UV absorption of sekothrixide at 276 nm in alkali solution. Further HMBC spectral analysis reveraled the remaining connectivity of the molecule from C-13 to C-21. The cross peaks between methyl proton signals (14-CH₃, 16-CH₃, 18-CH₃ and C-21) and oxygenated methines (C-15, C-17 and C-19), non-oxygenated methines (C-14, C-16 and C-18) or a methylene (C-20) enabled to determine the partial carbon skeleton from C-13 to C-21 (Fig. 1, II).

These partial structures I and II were connected through a non-equivalent methylene (12-H) at $\delta_{\rm H}$ 2.15 and 2.33, which showed ¹H-¹H couplings to 13-H and 11-H (Fig. 1, III).

The formation of a macrolide ring at C-13 in sekothrixide was indicated by the low field chemical shift of 13-H at 5.25 ppm. The geometry of the doublet bond (C-10 and C-11) was determined to be *E* based on the high field shift of 10-CH₃ ($\delta_{\rm C}$ 16.6)⁷⁾. Sekothrixide was thus demonstrated to be a 14-membered β -ketolactone with a side chain (Fig. 2). Its relative and absolute configurations remain to be established. Unlike known 14-membered

Fig. 2. Planar structure of sekothrixide.



macrolide antibiotics having a β -ketoester system such as rustmicin⁸⁾, neorustmicin⁹⁾ and pikromycin¹⁰⁾, sekothrixide is unique in possessing a polyoxygenated long side chain.

Sekothrixide exhibited neither antibacterial nor antifungal activity at a concentration of 1,000 μ g/ml. It inhibited in a dose dependent fashion, the growth of KB cells and its colchicine-resistant cells (KB-C2) with an IC₅₀ of 6.5 μ g/ml in EAGLE's minimum essential medium (EMEM) without colchicine. In the medium containing colchicine (1.5 μ g/ml), sekothrixide showed synergistic activity against KB-C2 with an IC₅₀ of 1.0 μ g/ml. Further studies on the stereochemistry and action mechanism of sekothrixide are in progress.

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References

- BRADLEY, G.; P. F. JURANKA & V. LING: Mechanism of multidrug resistance. Biochim. Biophys. Acta 948: 87~128, 1988
- 2) SHEN, D.; C. CARDARELLI, J. HWANG, M. CORNWELL, N. RICHERT, S. ISHII, I. PASTAN & M. M. GOTTESMAN: Multiple drug-resistant human KB carcinoma cells independently selected for high level resistant to colchicine, adriamycin, or vinblastin show changes in expression of specific proteins. J. Biol. Chem. 261: 7762~7770, 1986
- KIM, Y. J.; K. FURIHATA, S. YAMANAKA, R. FUDO & H. SETO: Isolation and structural elucidation of stipiamide, a new antibiotic effective to multidrugresistant cancer cells. J. Antibiotics 44: 553~556, 1991
- 4) BAX, A. & M. F. SUMMERS: ¹H and ¹³C assignments from sensitivity-enhanced detection of heteronuclear multiple-bond connectivity by 2D multiple quantum NMR. J. Am. Chem. Soc. 108: 2093~2094, 1986
- 5) SUMMERS, M. F.; L. G. MARZILLI & A. BAX: Complete ¹H and ¹³C assignments of coenzyme B₁₂ through the use of new two-dimensional NMR experiments. J. Am. Chem. Soc. 108: 4285~4294, 1986
- SETO, H.; K. FURIHATA & M. OHUCHI: Particular utility of the HMBC technique to polypropionate derived metabolites as examplified by erythromycin A. J. Antibiotics 41: 1158~1160, 1988
- BREITMAIER, E. & W. VOELTER (Ed.): Steric interactions. In ¹³C NMR spectroscopy. pp. 74~75, Verlag Chemie, 1978
- 8) TAKATSU, T.; H. NAKAYAMA, A. SHIMAZU, K. FURIHATA, K. IKEDA, K. FURIHATA, H. SETO & N. ŌTAKE: Rustmicin, a new macrolide antibiotic active against wheat stem rust fungus. J. Antibiotics 38: 1806~1809, 1985
- 9) NAKAYAMA, H.; T. HANAMURA, Y. ABE, A. SHIMAZU, K. FURIHATA, K. IKEDA, K. FURIHATA, H. SETO & N. ŌTAKE: Structures of neorustmicins B, C and D. New congeners of rustmicin and neorustmicin A. J. Antibiotics 39: 1016~1020, 1986
- MUXFELDT, H.; S. SHRADER, P. HANSEN & H. BROCKMANN: The structure of pikromycin. J. Am. Chem. Soc. 90: 4748~4749, 1968